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Practical synthesis of chiral ligands for catalytic enantioselective cyanosilylation of ketones

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Abstract—A practical and more environmentally benign synthetic route of chiral ligands that are useful for catalytic enantioselective cyanosilylation of ketones is described. A key step is the regioselective S_N^2 reaction to cyclic sulfate 12 with catechol derivatives. A range of chiral ligands containing electronically tuned catechols has become available by this new route. © 2002 Elsevier Science Ltd. All rights reserved.

1. Introduction

We recently developed a catalytic enantioselective cyanosilylation of ketones that affords chiral tertiary cyanohydrins from a wide range of ketone substrates.^{1,2} Both (R)- and (S)-cyanohydrins can be synthesized with generally high enantioselectivity by catalysts prepared from chiral ligand 1: 1-Ti (1:1) complex is (R)-selective and 1-Ln (Gd or Sm) (2:3) complex is (S)-selective (Scheme 1). The synthetic importance of chiral tertiary alcohols as building blocks for various pharmaceutically significant compounds (e.g. camptothecin, fostriecin, oxybutynin, pyruvate dehydrogenase kinase inhibitor, etc.)³ illustrates that 1 is an extremely useful chiral ligand.

1 was previously synthesized in high overall yield from commercially available tri-*O*-acetyl-D-glucal (7).⁴ A key step was the introduction of the catechol moiety (X in Scheme 1) by an aromatic substitution of the arene– chromium complex.⁵ Use of a stoichiometric amount of chromium, however, is environmentally problematic, especially in a large-scale synthesis. Furthermore, tuning the catechol moiety of 1 was very difficult using this synthetic route, because arene–chromium complexes containing electron-deficient aromatic groups are generally difficult to prepare. Introducing an electron-withdrawing group on the catechol moiety, however, is an attractive approach for improving the catalyst efficiency, because it increases the Lewis acidity of the metal. For example, benzoyl-containing ligand 2 synthesized by a laborious route produced significantly better results in reactions promoted by a Ti-complex.^{1b}





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Herein, we report a new and practical synthetic route of chiral ligands that is more suitable for large-scale synthesis (Scheme 2). The direct introduction of electrondeficient catechols allows for a facile synthesis of electronically tuned chiral ligands (2-6).

2. Practical synthetic route of chiral ligands

We attempted to introduce the key catechol via a regioselective $S_N 2$ attack of catechol derivatives to cyclic sulfate **12**. To do so required inversion of the 3-OH group of the D-glucose derivative **8** to the D-allose derivative **10**. Although there are several reports on the synthesis of **10**,⁶ none of them were satisfactory for our purpose. Therefore, we planned to develop an original method to synthesize **10** through an oxidation-stereoselective reduction sequence from **8**. Previous reports using related ketones indicated that the reduction with NaBH₄ should proceed mainly via an equatorial hydride attack to give the desired α -OH.⁷

Precursor ketone 9 was synthesized via Swern oxidation of $\mathbf{8}$,⁸ which was readily obtained from commercially available 7 through three steps. Reduction of 9 with NaBH₄ in MeOH, however, resulted in low stereoselectivity (8:10=1.5:1) and undesired 8 was obtained as the major isomer.⁹ After several attempts, reduction with L-selectride gave the desired 10 with complete selectivity in 93% yield from 8 (two steps).¹⁰ It is advantageous, especially for a large scale synthesis, that column chromatography was not necessary through 7-10, and all the purifications could be performed by recrystallization.¹¹ Deprotection of the benzylidene acetal, selective tosylation of the primary alcohol, and introduction of the phosphine oxide gave 11.¹² The key cyclic sulfate 12 was synthesized following the reported procedure.¹³ From 12, a variety of catechol moieties were introduced in an S_N2 manner with complete regioselectivity at the less hindered position (C-3).¹⁴ Deprotection of the methyl ether¹⁵ gave chiral ligands 1–6 [total 12 steps from 7 for 1, 2, 3, 5, and 6. 1: 17% yield (10 steps in 44% yield, previously). 2: 17% yield (13 steps in 10% yield, previously). 3: 19% yield. 4: 14% yield (total 15 steps). 5: 17% yield. 6: 25% yield.]. Ligands 3–6 cannot be synthesized by the former route using arene– chromium complexes.

In conclusion, we developed a practical and environmentally more benign (i.e. no chromium) synthetic route of chiral ligands that are very useful for catalytic enantioselective cyanosilylation of ketones. This synthetic route made it possible to readily prepare an array of chiral ligands containing different catechol moieties. The advantages of these new ligands will be discussed in a following paper.

3. Experimental

General procedure for the inversion of 3-OH (8–10): DMSO (0.21 mL, 2.96 mmol) was added dropwise to a solution of (COCl)₂ (0.13 mL, 1.49 mmol) in CH₂Cl₂ (5 mL) at -78°C. After 15 min, 8 (278 mg, 1.18 mmol) in CH₂Cl₂ (2 mL) was added dropwise for 10 min, and the resulting suspension was stirred at the same temperature for 1 h. Then, Et₃N (0.85 mL, 6.10 mmol) was added at -78°C and the dry ice-acetone bath was removed to allow the reaction temperature to reach room temperature. After 30 min, H₂O was added and the product was extracted with CHCl₃. Evaporation of the organic solvent gave crude ketone 9, which can be used for the next step without purification. L-Selectride (1.0 mol L⁻¹ in THF, 467 μ L, 0.47 mmol) was added to a solution of ketone 9 (0.44 mmol) in THF (4 mL) at -78°C. After 50 min, 30% H₂O₂ aq. (0.32 mL) was added at the same temperature and the mixture was stirred at 4°C for 20 min. The mixture was then slowly poured into sat. $Na_2S_2O_3$ aq. and the aqueous layer was



Scheme 2. *Reagents and conditions*: (a) Pd/C, H₂, MeOH, rt; (b) NaOMe (0.25 equiv.), MeOH, rt; (c) PhCH(OMe)₂ (1.2 equiv.), *p*-TsOH·H₂O (0.3 equiv.), toluene, reflux, 57% (three steps); (d) DMSO (2.5 equiv.), (COCl)₂ (1.3 equiv.), Et₃N (5 equiv.), CH₂Cl₂, -78°C to rt; (e) L-Selectride (1.03 equiv.), THF, -78°C, 93% (two steps); (f) Pd/C, H₂, MeOH–AcOH (1:1), rt, 92%; (g) TsCl (1.1 equiv.), pyridine, 78%; (h) Ph₂PK (2.5 equiv.), THF, 0°C; H₂O₂, 94% (Ar=Ph), [Ar=*p*-CH₃C₆H₄: MOMCl (2.5 equiv.), 'Pr₂NEt (3 equiv.), CH₂Cl₂, reflux, 72%; Ar₂PLi (2 equiv.), THF, 0°C; H₂O₂; cat. HCl aq.–MeOH, reflux, 85% (three steps)]; (i) SOCl₂ (1.2 equiv.), CH₂Cl₂; (j) RuCl₃·3H₂O (1 mol%), NaIO₄ (1.5 equiv.), CCl₄–CH₃CN–H₂O, 90% (Ar=Ph), 98% (Ar=*p*-CH₃C₆H₄) (two steps); (k) catechol derivatives (1.3 equiv.), K₂CO₃ (2 equiv.), DMF; 20% H₂SO₄ aq., Et₂O/CHCl₃, 65–86% (two steps); (l) EtSH, AlCl₃, CH₂Cl₂, 82–100% (two steps).

extracted with Et₂O. The combined organic layer was washed with brine and dried over Na_2SO_4 . Evaporation and purification by SiO₂ column chromatography (eluent: AcOEt/hexane=1/2) gave pure **10** in 93% yield as white powder. In a large scale synthesis, the crude product was purified by recrystallization (AcOEt–hexane).

General procedure for the introduction of the phosphine oxide (11): A solution of the tosylate (8.57 g, 28.3 mmol) in THF (30 mL) was added to a commercially available Ph₂PK solution in THF (0.5 M, 142 mL, 71.0 mmol) over 10 min in an ice bath. After stirring for 30 min at the same temperature, sat. NH₄Cl aq. was added, followed by the addition of 30% H₂O₂ aq. (16 mL) under vigorous stirring. The mixture was then poured slowly into sat. $Na_2S_2O_3$ aq. in an ice bath. Evaporation of THF, extraction of the aqueous layer with CHCl₃, and concentration gave a crude oil that was purified by SiO₂ column chromatography (eluent: $MeOH/CHCl_3 = 1/30$). The resulting solid was recrystallized from CHCl₃-Et₂O to give white powder in 94% yield. The corresponding diol containing di-(ptolyl)phosphine oxide was synthesized from the MOM protected tosylate in a similar way using (p-tolyl)₂PLi prepared following the reported procedure.¹⁶

General procedure for the cyclic sulfate synthesis (12): SOCl₂ (0.27 mL, 3.70 mmol) was added to a solution of diol 11 (1.03 g, 3.10 mmol) in CH₂Cl₂ (20 mL), and the mixture was stirred for 30 min at room temperature. Then, the reaction mixture was concentrated, followed by co-evaporation with toluene two times. The resulting crude cyclic sulfite was dissolved in a CCl₄ (15 mL)/CH₃CN (15 mL)/H₂O (30 mL), and NaIO₄ (994 mg, 4.65 mmol) and RuCl₃·nH₂O (7.3 mg, 1 mol%) were added in an ice bath. After stirring vigorously for 1 h, H₂O was added and the aqueous layer was extracted with AcOEt. The combined organic layer was washed with sat. NaCl aq., followed by stirring with PrOH (5 mL) at room temperature for 1 h. Drying with Na₂SO₄, filtration through celite, and concentration gave a crude powder, which was recrystallized from AcOEt-Et₂O to give 12 as fine needles (1.10 g, 90%).

General procedure for the $S_N 2$ reaction by catechol derivatives: K_2CO_3 (1.32 g, 9.56 mmol) and guaiacol (0.68 mL, 6.21 mmol) were added to a solution of 12 (1.86 g, 4.78 mmol) in DMF (10 mL) and the mixture was stirred for 14 h at room temperature. After 12 had disappeared on TLC, 20% H₂SO₄ aq. (100 mL) and Et₂O (50 mL)–CHCl₃ (50 mL) were added and the mixture was vigorously stirred for 12 h. The aqueous layer was extracted with CHCl₃ and the combined organic layer was washed with H₂O and sat. NaCl aq. Drying over Na₂SO₄, followed by concentration, gave a crude oil that was purified by SiO₂ column chromatography (eluent: AcOEt/hexane=1/1 to 100/0). The methylated ligand was obtained as a white solid (65%).

4. Selected spectral data of new ligands

3: ¹H NMR (500 MHz, CDCl₃) δ 1.94 (dddd, J=5.1, 12.9, 12.9, 12.9 Hz, 1H), 2.09 (m, 1H), 2.67 (ddd, J=9.7, 15.0, 15.0 Hz, 1H), 2.85 (ddd, J=2.5, 9.7, 15.0 Hz, 1H), 3.24 (ddd, J = 1.8, 12.3, 12.9 Hz, 1H), 3.33 (m, 1H), 3.50 (ddd, J=5.1, 8.6, 12.9 Hz, 1H), 3.69 (brt, J = 8.6 Hz, 1H), 3.91 (m, 1H), 6.77 (ddd, J = 8.2, 10.7, 19.8 Hz, 1H), 7.53 (m, 6H), 7.76 (m, 4H), 7.85 (s, 1H), 9.16 (d, J=1.2 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 31.56, 36.04 (d, J=69 Hz), 65.39, 74.70 (d, J=3 Hz), 75.93, 85.57, 105.58 (d, J=21 Hz), 110.93 (d, J=18Hz), 129.03 (d, J = 12 Hz), 129.07 (d, J = 12 Hz), 130.20 (d, J = 101 Hz), 130.71 (d, J = 10 Hz), 131.17 (d, J = 10Hz), 131.62 (d, J=100 Hz), 132.58 (d, J=4 Hz), 132.61 (d, J=3 Hz), 141.14 (dd, J=3, 7 Hz), 142.60 (dd, J=14, 239 Hz), 146.58 (dd, J=3, 10 Hz), 147.37 (dd, J = 13, 243 Hz). ³¹P NMR (202 MHz, CDCl₃) δ 33.3.

4: ¹H NMR (500 MHz, CDCl₃) δ 1.93 (dddd, J=5.2, 12.8, 12.8, 12.8 Hz, 1H), 2.08 (m, 1H), 2.40 (s, 3H), 2.41 (s, 3H), 2.63 (ddd, J=10.1, 15.3, 15.3 Hz, 1H), 2.79 (ddd, J=2.8, 9.8, 15.3 Hz, 1H), 3.23 (ddd, J=2.1, 11.9,12.8 Hz, 1H), 3.30 (m, 1H), 3.49 (ddd, J=5.2, 8.9, 11.3 Hz, 1H), 3.67 (brt, J = 8.9 Hz), 3.91 (m, 1H), 6.74 (dd, J=7.9, 11.6 Hz, 1H), 6.78 (dd, J=8.3, 10.7 Hz, 1H), 7.29 (dd, J=2.8, 8.3 Hz, 2H), 7.32 (dd, J=2.8, 8.3 Hz, 2H), 7.58 (dd, J=8.3, 11.9 Hz, 2H), 7.62 (dd, J=8.3, 11.9 Hz, 2H), 7.97 (s, 1H), 9.24 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 21.71, 21.73, 31.55, 36.42 (d, J=69Hz), 65.38, 74.72 (d, J=3 Hz), 76.01, 85.66, 105.58 (d, J = 22 Hz), 110.99 (d, J = 20 Hz), 126.80 (d, J = 103 Hz), 128.36 (d, J=108 Hz), 129.74 (d, J=12 Hz), 129.83 (d, J=13 Hz), 130.71 (d, J=10 Hz), 131.19 (d, J=9 Hz), 141.17 (dd, J=3, 8 Hz), 142.55 (dd, J=14, 239 Hz), 143.19 (d, J=2 Hz), 143.20 (d, J=2 Hz), 146.67 (dd, J=2, 10 Hz), 147.38 (dd, J=13, 243 Hz). ³¹P NMR (202 MHz, CDCl₃) δ 33.7.

5: ¹H NMR (500 MHz, CDCl₃) δ 1.99 (dddd, J=4.9, 12.8, 12.8, 12.8 Hz, 1H), 2.23 (m, 1H), 2.73 (ddd, J=9.5, 15.0, 15.0 Hz, 1H), 2.84 (ddd, J=3.1, 9.5, 15.0 Hz, 1H), 3.24 (ddd, J=1.9, 12.2, 12.8 Hz, 1H), 3.39 (m, 1H), 3.71 (ddd, J=4.9, 8.9, 11.0 Hz, 1H), 3.79 (t like, J=8.9 Hz, 1H), 3.91 (m, 1H), 7.25 (brt, J=7.0 Hz, 1H), 7.30 (s, 1H), 7.33 (brt, J=7.0 Hz, 1H), 7.36 (s, 1H), 7.5–7.66 (m, 8H), 7.77 (m, 4H), 8.98 (brs, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 31.42, 35.95 (d, J=69 Hz), 65.49, 74.82 (d, J=4 Hz), 76.05, 84.16, 111.55, 117.54, 123.41, 125.10, 126.32, 126.86, 128.36, 128.98 (d, J=12 Hz), 129.01 (d, J=12 Hz), 130.60 (d, J=101 Hz), 130.78 (d, J=10 Hz), 131.23 (d, J=9 Hz), 131.97 (d, J=103 Hz), 132.04, 132.44 (d, J=2 Hz), 146.88, 149.15. ³¹P NMR (202 MHz, CDCl₃) δ 33.3.

6: ¹H NMR (500 MHz, CDCl₃) δ 2.11 (dddd, J=4.9, 12.5, 12.5, 12.5 Hz, 1H), 2.26 (m, 1H), 2.69 (ddd, J=10.1, 15.3, 15.3 Hz, 1H), 2.85 (m, 1H), 3.21 (brt, J=12.5 Hz, 1H), 3.36 (m, 1H), 3.70 (ddd, J=4.9, 8.9, 11.3 Hz, 1H), 3.81 (t like, 8.9 Hz, 1H), 3.90 (m, 1H), 7.49–7.63 (m, 6H), 7.72 (dd, J=7.3, 12.2 Hz, 2H), 7.78 (dd, J=7.3, 11.9 Hz, 2H), 8.47 (s, 1H), 10.9 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 32.93, 36.02 (d, J=69 Hz),

65.39, 74.85 (d, J=3 Hz), 75.67, 87.03, 120.58, 121.92, 126.81, 128.73, 129.06 (d, J=11 Hz), 129.10 (d, J=12 Hz), 129.90 (d, J=101 Hz), 130.62 (d, J=10 Hz), 131.20 (d, J=10 Hz), 131.57 (d, J=100 Hz), 132.65 (d, J=3 Hz), 142.78, 147.77. ³¹P NMR (202 MHz, CDCl₃) δ 33.4.

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